

Supplementary information S1 (table) | **Survey of the regulation of bacterial antibiotics.**

We report cases where there is a strong candidate for the regulator of the antibiotic. The examples are divided into categories based upon the condition and regulator associated with the induction of the antibiotic. Note that the role of any one regulator is typically evidenced by gene deletion or over-expression, which are both potentially subject to confounding pleiotropic effects <sup>1</sup>. For toxins that are activated towards stationary phase in batch culture, inference of the regulators is particularly important and we use this to distinguish between stress response regulation and quorum associated regulation. Sometimes both are important e.g. bacilysin. We include two interesting sets of examples where regulators are not known, which are halocins (stationary phase associated toxins in Archea) and the pH based quorum-like regulation of lactic acid bacteria. However, examples where the regulator is not known are excluded from the statistics in the paper. Finally, the table does not include cases where a specific nutrient source affects toxin production or where certain non-stressful temperatures produced more induction than others as these are often difficult to interpret (see main text).

Condition	Species	Regulator	Antibiotic <sup>c,d</sup>	Effect of condition	Bacteriocin?	Notes and references
Stringent Response	<i>Bacillus subtilis</i>	ppGpp	Bacilysin (1)	+	N	2
	<i>Escherichia coli</i>	ppGpp	Colicin K (2)	+	Y	3
	<i>Escherichia coli</i>	ppGpp	Microcin J (3)	+	Y	4
	<i>Pseudomonas aeruginosa</i>	ppGpp	Pyocyanin (4)	-	N	5, ppGpp reduces toxin but pushes release later: stationary phase regulation?
	<i>Streptomyces coelicolor</i>	ppGpp	Actinorhodin, CDA (calcium-dependent antibiotic), Undecylprodigiosin (7)	+	N	6-8
	<i>Streptomyces hygroscopicus</i>	ppGpp?	Bialaphos (8)	+	N	9, inference of ppGpp role is correlative.
	<i>Streptomyces antibioticus</i>	ppGpp	Actinomycin (9)	+	N	10
	<i>Streptomyces lavendulae</i>	ppGpp	Formycin (10)	+	N	11
	<i>Streptomyces griseus</i>	ppGpp	Streptomycin (11)	+	N	12
	<i>Streptomyces tendae</i>	ppGpp	Nikkomycin (12)	+	N	13
	<i>Streptomyces clavuligerus</i>	ppGpp	Cephameycin C	+	N	14
	<i>Streptomyces clavuligerus</i>	ppGpp	Cephameycin C (13)	-	N	15, conflicts with 14, we use 15 to be conservative (also, see 16).
	General stress responses and other nutrient	<i>Bacillus subtilis</i>	σH	Subtilin (14)	+	Y
<i>Bacillus subtilis</i>		Spo0A	Sporulation killing factor (15)	+	Y	19, role in fratricide rather

limitation responses <sup>a</sup>						than competition?
	<i>Bacillus subtilis</i>	AbrB/ Spo0A	Subtilisin (16)	+	Y	20
	<i>Bacillus subtilis</i>	AbrB/ Spo0A	TasA (17)	+	N	21
	<i>Bacillus subtilis</i>	CodY	Bacilysin (18)	+	N	2
	<i>Escherichia coli</i>	IHF	Microcin B (19)	+	Y	4, 22
	<i>Escherichia coli</i>	σS	Microcin C (20)	+	Y	4
	<i>Escherichia coli</i>	IHF/LRP	Microcin J (21)	+	Y	4
	Various <i>Halobacteriaceae</i> (Archaea)	Unknown <sup>b</sup>	Halocin A4, C8, G1, H2, H3, H4, H5, H6, H7, R1, S8	+	Y	23, role for quorum regulation?
	<i>Haloferax mediterranei</i> M2a	Unknown <sup>b</sup>	H1	-	Y	23
	<i>Pseudomonas fluorescens</i>	σS	Pyrrrolnitrin (22)	+	N	24
	<i>Pseudomonas fluorescens</i>	σS	Pyoluteorin and 2,4-diacetylphloroglucinol (24)	-	N	24
	<i>Pseudomonas aeruginosa</i>	σS	Pyocyanin (25)	-	N	25
	<i>Streptococcus pneumoniae</i>	CodY	Putative bacteriocin (26)	+	Y	26
	<i>Streptococcus gordonii</i>	CcpA	H2O2 (27)	+	N	27
	<i>Streptomyces coelicolor</i>	σB	Undecylprodigiosin (28)	+	N	28,29
	<i>Streptomyces coelicolor</i>	σB	Actinorhodin (29)	-	N	28,29
	<i>Streptomyces coelicolor</i>	σL	Actinorhodin (30)	+	N	29
Envelope stress	<i>Bacillus subtilis</i>	σW	Probable bacteriocin (31)	+	Y	30
	<i>Streptomyces coelicolor</i>	σE	Actinorhodin, CDA, undecylprodigiosin (34)	+	N	6
DNA damage	<i>Clostridium perfringens</i>	UviA	bacteriocin BCN5(35)	+	Y	31,32
	<i>Escherichia coli</i>	LexA /RecA	Colicin A, B, D157, E1, E2, E3, E6, E7, Js, N, S4, U,Y, Ia, Ib, 5, 10, K (53)	+	Y	33-36
	<i>Hafnia alvei</i>	LexA	Alveicins A, B (55)	+	Y	37
	<i>Klebsiella pneumoniae</i>	LexA	Klebicins B, C (57)	+	Y	38,39
	<i>Klebsiella oxytoca</i>	LexA	Klebicins B, D (58)	+	Y	38,39
	<i>Pectobacterium carotovorum</i>	RdgB/RecA	Carotovoricin (Ctv) and pectin lyase (Pnl) (60)	+	Y	40
	<i>Pectobacterium carotovorum</i>	RdgB/RecA	Carocin D (61)	+	Y	41
	<i>Pseudomonas aeruginosa</i>	RecA	Pyocin R1, R2, R3, F1, F2, F3, S1, S2, S3, S4, S5, AP41 (73)	+	Y	42,43
	<i>Pseudomonas putida</i>	RecA	Lectin-like putidacin A (74)	+	Y	44

	<i>Serratia marcescens</i>	LexA/ RegC	Bacteriocin 28B (75)	+	Y	45
	<i>Yersinia pestis</i>	LexA	Pesticin (76)	+	Y	46
<b>Oxidative stress</b>	<i>Escherichia coli</i>	LexA/ RecA?	Colicin	+	Y	47
	<i>Pseudomonas aeruginosa</i>	RecA?	Pyocin S2, S3, S5, R2, F2	+	Y	48
	<i>Streptococcus mutans</i>	VicRK	Mutacin I, Mutacin IV, V and VI (80)	+	Y	49-52
	<i>Streptomyces coelicolor</i>	SoxR	Actinorhodin (81)	+	Y	53
<b>Heat stress</b>	<i>Escherichia coli</i>	σ32	Colicin A (82)	+	Y	54
	<i>Pseudomonas fluorescens</i>	Lon	pyoluteorin (83)	-	N	55
<b>Osmotic stress</b>	<i>Escherichia coli</i>	OmpR	Microcin B (84)	+	Y	4
	<i>Streptomyces coelicolor</i>	σB/ <i>OsaB</i>	Actinorhodin (85)	-	N	56
<b>Quorum sensing</b>	<i>Bacillus subtilis</i>	Spo0K	Bacilysin	+	N	57
	<i>Burkholderia Thailandensis</i>	LuxI/LuxR	Unnamed Antibiotic	+	N	58
	<i>Chromobacterium violaceum</i>	LuxI/LuxR	Violacein	+	N	59
	<i>Enterococcus faecium CTC492</i>	EntK/EntR	Enterocin A, enterocin B	+	Y	60
	<i>Erwinia carotovora</i>	LuxI/LuxR	Carbapenem	+	N	61
	<i>Streptomyces coelicolor</i> , <i>S. griseus</i> and others	γ-butyrolactone receptor systems	Various antibiotics	+	N	62,63
	<i>Lactobacillus plantarum C11</i>	<i>plnABCD</i>	Multiple bacteriocins	+	Y	64
	<i>Lactobacillus Sake</i>	<i>sppIPKR</i>	Sakacin P	+	Y	65,66
	<i>Streptococcus mutans</i>	ComCDE	Mutacin IV	+	Y	67
	<i>Streptococcus mutans</i>	LuxS (AI2), CiaXRH	Mutacin I	+	Y	68,69
	<i>Streptococcus pneumoniae</i>	BlpSRH	Blp bacteriocins	+	Y	70
	<i>Streptococcus pyogenes</i>	SalKR	SalA1	+	Y	71
	<i>Streptococcus thermophilus</i>	BlpHSt-BlpRSt	Blp bacteriocins	+	Y	72
<b>Toxin autoinduction</b>	<i>Bacillus sp. strain HIL Y-85,54728</i>	MrsR2/MrsK 2	Mersacidin	+	Y	73
	<i>Bacillus subtilis</i>	SpaK	Subtilin	+	N	17
	<i>Carnobacterium piscicola LV17B</i>	<i>CbnK</i> , <i>CbnR</i>	Carnobacteriocin B2 carnobacteriocin BM1	+	Y	74
	<i>Lactococcus lactis</i>	NisK	Nisin	+	Y	75

	<i>Streptococcus salivarius</i> 20P3	SalKR two component CylR1, CylR2	Salivaricin A (Sala)	+	Y	71
	<i>Enterococcus faecalis</i>		Cytolysin	+	N	76
<b>Quorum: other</b>	<i>Pseudomonas aeruginosa</i>	<i>nagR</i> operon	Pyocyanin	+	N	77, Peptidoglycan is inducer.
<b>Low pH</b>	<i>Lactococcus lactis</i>	Unknown	Nisin	+	Y	78
	<i>Lactobacillus Sake</i>	Unknown	Lactocin S	+	Y	79
	<i>Pediococcus acidilactici</i>	Unknown	Pediocin	+	Y	78
	<i>Lactococcus lactis</i>	Unknown	Lacticin 481	+	Y	80
<b>Constitutive</b>	<i>Lactococcus lactis</i>		Lacticin 3147		Y	81
	<i>Enterococcus faecium</i> BFE 900		enterocin B		Y	82

**Footnotes:**

- a) Often also known as “stationary phase regulators” but can activate under continuous exponential or linear growth when there is a continuous supply of nutrients<sup>83</sup>, particularly in biofilms where cell density is high<sup>84</sup>.
- b) These are not included in the count of links as the regulators are unknown.
- c) Antibiotics that are regulated by more than one condition are coloured by species.
- d) The numbers in parenthesis after the antibiotic count up the number of distinct links between a stress response regulator and an antibiotic.

**References**

1. Bibb, M.J. Regulation of secondary metabolism in streptomycetes. *Current Opinion in Microbiology* **8**, 208-215 (2005).
2. Inaoka, T., Takahashi, K., Ohnishi-Kameyama, M., Yoshida, M. & Ochi, K. Guanine nucleotides guanosine 5'-diphosphate 3'-diphosphate and GTP cooperatively regulate the production of an antibiotic bacilysin in *Bacillus subtilis*. *Journal of Biological Chemistry* **278**, 2169-2176 (2003).
3. Kuhar, I. & Žgur-Bertok, D. Transcription Regulation of the Colicin Kcka Gene Reveals Induction of Colicin Synthesis by Differential Responses to Environmental Signals. *Journal of Bacteriology* **181**, 7373-7380 (1999).
4. Moreno, F., González-Pastor, J.E., Baquero, M.R. & Bravo, D. The regulation of microcin B, C and J operons. *Biochimie* **84**, 521-529 (2002).
5. Erickson, D.L., Lines, J.L., Pesci, E.C., Venturi, V. & Storey, D.G. *Pseudomonas aeruginosa* *relA* Contributes to Virulence in *Drosophila melanogaster*. *Infection and Immunity* **72**, 5638-5645 (2004).
6. Hesketh, A. et al. Genome-wide dynamics of a bacterial response to antibiotics that target the cell envelope. *BMC Genomics* **12**, 226 (2011).
7. Hesketh, A., Chen, W.J., Ryding, J., Chang, S. & Bibb, M. The global role of ppGpp synthesis in morphological differentiation and antibiotic production in *Streptomyces coelicolor* A3 (2). *Genome Biol* **8**, R161 (2007).
8. Strauch, E., Takano, E., Baylts, H. & Bibb, M. The stringent response in *Streptomyces coelicolor* A3 (2). *Molecular Microbiology* **5**, 289-298 (1991).
9. Holt, T.G. et al. Global changes in gene expression related to antibiotic synthesis in *Streptomyces hygroscopicus*. *Molecular Microbiology* **6**, 969-980 (1992).
10. Hoyt, S. & Jones, G.H. *relA* is required for actinomycin production in *Streptomyces antibioticus*. *Journal of Bacteriology* **181**, 3824-3829 (1999).

11. Ochi, K. Occurrence of the stringent response in *Streptomyces* sp. and its significance for the initiation of morphological and physiological differentiation. *Journal of general microbiology* **132**, 2621-2631 (1986).
12. Ochi, K. Metabolic initiation of differentiation and secondary metabolism by *Streptomyces griseus*: significance of the stringent response (ppGpp) and GTP content in relation to A factor. *Journal of Bacteriology* **169**, 3608-3616 (1987).
13. Pfefferle, U., Ochi, K. & Fiedler, H.P. The Stringent Response and the Induction of Nikkomycin Production in *Streptomyces tendae*. *Actinomycetologica* **9**, 118-123 (1995).
14. Jin, W. et al. Cephamycin C production is regulated by *relA* and *rsh* genes in *Streptomyces clavuligerus* ATCC27064. *Journal of Biotechnology* **114**, 81-87 (2004).
15. Gomez-Escribano, J.P., Martín, J.F., Hesketh, A., Bibb, M. & Liras, P. *Streptomyces clavuligerus* *relA*-null mutants overproduce clavulanic acid and cephamycin C: negative regulation of secondary metabolism by (p) ppGpp. *Microbiology* **154**, 744-755 (2008).
16. Liras, P., Gomez-Escribano, J.P. & Santamarta, I. Regulatory mechanisms controlling antibiotic production in *Streptomyces clavuligerus*. *Journal of Industrial Microbiology & Biotechnology* **35**, 667-676 (2008).
17. Kleerebezem, M. Quorum sensing control of lantibiotic production; nisin and subtilin autoregulate their own biosynthesis. *Peptides* **25**, 1405-1414 (2004).
18. Stein, T. et al. Dual control of subtilin biosynthesis and immunity in *Bacillus subtilis*. *Molecular Microbiology* **44**, 403-416 (2002).
19. Gonzalez-Pastor, J.E., Hobbs, E.C. & Losick, R. Cannibalism by sporulating bacteria. *Science* **301**, 510-513 (2003).
20. Nakano, M.M., Zheng, G. & Zuber, P. Dual Control of *sbo*-*alb* Operon Expression by the Spo0 and ResDE Systems of Signal Transduction under Anaerobic Conditions in *Bacillus subtilis*. *Journal of Bacteriology* **182**, 3274-3277 (2000).
21. Stöver, A.G. & Driks, A. Regulation of Synthesis of the *Bacillus subtilis* Transition-Phase, Spore-Associated Antibacterial Protein TasA. *Journal of Bacteriology* **181**, 5476-5481 (1999).
22. Navarro Llorens, J.M., Tormo, A. & Martínez-García, E. Stationary phase in gram-negative bacteria. *FEMS Microbiology Reviews* **34**, 476-495 (2010).
23. Shand, R.F. & Leyva, K.J. Peptide and Protein Antibiotics from the Domain Archaea: Halocins and Sulfolobocins  
Bacteriocins. (eds. Riley, M.A. & Chavan, M.A.) 93-109 (Springer Berlin Heidelberg, 2007).
24. Sarniguet, A., Kraus, J., Henkels, M.D., Muehlchen, A.M. & Loper, J.E. The sigma factor sigma s affects antibiotic production and biological control activity of *Pseudomonas fluorescens* Pf-5. *Proceedings of the National Academy of Sciences* **92**, 12255 (1995).
25. Schuster, M., Hawkins, A.C., Harwood, C.S. & Greenberg, E.P. The *Pseudomonas aeruginosa* RpoS regulon and its relationship to quorum sensing. *Molecular Microbiology* **51**, 973-985 (2004).
26. Hendriksen, W.T. et al. CodY of *Streptococcus pneumoniae*: link between nutritional gene regulation and colonization. *Journal of Bacteriology* **190**, 590-601 (2008).
27. Zheng, L., Itzek, A., Chen, Z. & Kreth, J. Environmental influences on competitive hydrogen peroxide production in *Streptococcus gordonii*. *Applied and Environmental Microbiology* **77**, 4318-4328 (2011).
28. Cho, Y.H., Lee, E.J., Ahn, B.E. & Roe, J.H. SigB, an RNA polymerase sigma factor required for osmoprotection and proper differentiation of *Streptomyces coelicolor*. *Molecular Microbiology* **42**, 205-214 (2001).

29. Lee, E.J. et al. A master regulator  $\sigma^B$  governs osmotic and oxidative response as well as differentiation via a network of sigma factors in *Streptomyces coelicolor*. *Molecular Microbiology* **57**, 1252-1264 (2005).
30. Cao, M. et al. Defining the *Bacillus subtilis* [sigma] W regulon: a comparative analysis of promoter consensus search, run-off transcription/microarray analysis (ROMA), and transcriptional profiling approaches. *Journal of Molecular Biology* **316**, 443-457 (2002).
31. Garnier, T. & Cole, S.T. Characterization of a bacteriocinogenic plasmid from *Clostridium perfringens* and molecular genetic analysis of the bacteriocin-encoding gene. *Journal of Bacteriology* **168**, 1189-1196 (1986).
32. Dupuy, B., Mani, N., Katayama, S. & Sonenshein, A.L. Transcription activation of a UV-inducible *Clostridium perfringens* bacteriocin gene by a novel  $\sigma$  factor. *Molecular Microbiology* **55**, 1196-1206 (2005).
33. Walker, D. et al. Transcriptional profiling of colicin-induced cell death of *Escherichia coli* MG1655 identifies potential mechanisms by which bacteriocins promote bacterial diversity. *Journal of Bacteriology* **186**, 866 (2004).
34. Jerman, B., Butala, M. & Žgur-Bertok, D. Sublethal concentrations of ciprofloxacin induce bacteriocin synthesis in *Escherichia coli*. *Antimicrobial Agents and Chemotherapy* **49**, 3087-3090 (2005).
35. Majeed, H., Gillor, O., Kerr, B. & Riley, M.A. Competitive interactions in *Escherichia coli* populations: the role of bacteriocins. *The ISME Journal* **5**, 71-81 (2010).
36. Chavan, M. & Riley, M. Molecular Evolution of Bacteriocins in Gram-Negative Bacteria. in *Bacteriocins. Ecology and Evolution* (eds. Riley, M.A. & Chavan, M.A.) 19-43 (Springer Berlin Heidelberg, 2007).
37. Wertz, J.E. & Riley, M.A. Chimeric nature of two plasmids of *Hafnia alvei* encoding the bacteriocins alveicins A and B. *Journal of Bacteriology* **186**, 1598-1605 (2004).
38. Chavan, M., Rafi, H., Wertz, J., Goldstone, C. & Riley, M.A. Phage associated bacteriocins reveal a novel mechanism for bacteriocin diversification in *Klebsiella*. *Journal of Molecular Evolution* **60**, 546-556 (2005).
39. Gillor, O., Vriezen, J.A.C. & Riley, M.A. The role of SOS boxes in enteric bacteriocin regulation. *Microbiology* **154**, 1783-1792 (2008).
40. Yamada, K., Kaneko, J., Kamio, Y. & Itoh, Y. Binding sequences for RdgB, a DNA damage-responsive transcriptional activator, and temperature-dependent expression of bacteriocin and pectin lyase genes in *Pectobacterium carotovorum* subsp. *carotovorum*. *Applied and Environmental Microbiology* **74**, 6017-6025 (2008).
41. Roh, E. et al. Characterization of a New Bacteriocin, Carocin D, from *Pectobacterium carotovorum* subsp. *carotovorum* Pcc21. *Applied and Environmental Microbiology* **76**, 7541-7549 (2010).
42. Matsui, H., Sano, Y., Ishihara, H. & Shinomiya, T. Regulation of pyocin genes in *Pseudomonas aeruginosa* by positive (prtN) and negative (prtR) regulatory genes. *Journal of Bacteriology* **175**, 1257-1263 (1993).
43. Michel-Briand, Y. & Baysse, C. The pyocins of *Pseudomonas aeruginosa*. *Biochimie* **84**, 499-510 (2002).
44. De Los Santos, P.E., Parret, A.H.A. & De Mot, R. Stress-related *Pseudomonas* genes involved in production of bacteriocin LlpA. *FEMS Microbiology Letters* **244**, 243-250 (2005).
45. Ferrer, S., Viejo, M.B., Guasch, J.F., Enfedaque, J. & Regué, M. Genetic evidence for an activator required for induction of colicin-like bacteriocin 28b production in *Serratia marcescens* by DNA-damaging agents. *Journal of Bacteriology* **178**, 951-960 (1996).
46. Rakin, A., Boolgakowa, E. & Heesemann, J. Structural and functional organization of the *Yersinia pestis* bacteriocin pesticin gene cluster. *Microbiology* **142**, 3415-3424 (1996).

47. Šmarda, J. & Čermák, V. Induction of the formation of a coli-bacterio-phage and colicin by hydroperoxide. *Cellular and Molecular Life Sciences* **18**, 271-273 (1962).
48. Chang, W., Small, D.A., Toghrol, F. & Bentley, W.E. Microarray analysis of *Pseudomonas aeruginosa* reveals induction of pyocin genes in response to hydrogen peroxide. *BMC Genomics* **6**, 1-14 (2005).
49. Deng, D.M., Liu, M.J., ten Cate, J.M. & Crielaard, W. The VicRK System of *Streptococcus mutans* Responds to Oxidative Stress. *Journal of Dental Research* **86**, 606-610 (2007).
50. Senadheera, D.B. et al. Regulation of Bacteriocin Production and Cell Death by the VicRK Signaling System in *Streptococcus mutans*. *Journal of Bacteriology* **194**, 1307-1316 (2012).
51. Kreth, J., Merritt, J., Shi, W. & Qi, F. Competition and Coexistence between *Streptococcus mutans* and *Streptococcus sanguinis* in the Dental Biofilm. *Journal of Bacteriology* **187**, 7193-7203 (2005).
52. Liu, J., Wu, C., Huang, I.-H., Merritt, J. & Qi, F. Differential response of *Streptococcus mutans* towards friend and foe in mixed-species cultures. *Microbiology* **157**, 2433-2444 (2011).
53. Shin, J.-H., Singh, A.K., Cheon, D.-J. & Roe, J.-H. Activation of the SoxR Regulon in *Streptomyces coelicolor* by the Extracellular Form of the Pigmented Antibiotic Actinorhodin. *Journal of Bacteriology* **193**, 75-81 (2011).
54. Cavard, D. Effects of temperature and of heat shock on the expression and action of the colicin A lysis protein. *Journal of Bacteriology* **177**, 5189 (1995).
55. Whistler, C.A., Stockwell, V.O. & Loper, J.E. Lon Protease Influences Antibiotic Production and UV Tolerance of *Pseudomonas fluorescens* Pf-5. *Applied and Environmental Microbiology* **66**, 2718-2725 (2000).
56. Martínez, L.F. et al. Osmoregulation in *Streptomyces coelicolor*: modulation of SigB activity by OsaC. *Molecular Microbiology* **71**, 1250-1262 (2009).
57. Yazgan, A., Özcengiz, G. & Marahiel, M.A. Tn10 insertional mutations of *Bacillus subtilis* that block the biosynthesis of bacilysin. *Biochimica et Biophysica Acta (BBA) - Gene Structure and Expression* **1518**, 87-94 (2001).
58. Duerkop, B.A. et al. Quorum-sensing control of antibiotic synthesis in *Burkholderia thailandensis*. *Journal of Bacteriology* **191**, 3909-3918 (2009).
59. McClean, K.H. et al. Quorum sensing and *Chromobacterium violaceum*: exploitation of violacein production and inhibition for the detection of N-acylhomoserine lactones. *Microbiology* **143**, 3703-3711 (1997).
60. Nilsen, T., Nes, I.F. & Holo, H. An Exported Inducer Peptide Regulates Bacteriocin Production in *Enterococcus faecium* CTC492. *Journal of Bacteriology* **180**, 1848-1854 (1998).
61. Bainton, N. et al. N-(3-oxohexanoyl)-L-homoserine lactone regulates carbapenem antibiotic production in *Erwinia carotovora*. *Biochemical Journal* **288**, 997 (1992).
62. Takano, E.  $\gamma$ -Butyrolactones: *Streptomyces* signalling molecules regulating antibiotic production and differentiation. *Current Opinion in Microbiology* **9**, 287-294 (2006).
63. Khokhlov, A.S. et al. The A-factor, responsible for streptomycin biosynthesis by mutant strains of *Actinomyces streptomycini*. *Dokl Akad Nauk SSSR* **177**, 232-235 (1967).
64. Diep, D.B., Håvarstein, L.S. & Nes, I.F. Characterization of the locus responsible for the bacteriocin production in *Lactobacillus plantarum* C11. *Journal of Bacteriology* **178**, 4472-83 (1996).

65. Eijsink, V., Brurberg, M.B., Middelhoven, P.H. & Nes, I.F. Induction of bacteriocin production in *Lactobacillus sake* by a secreted peptide. *Journal of Bacteriology* **178**, 2232-2237 (1996).
66. Risøen, P.A., Brurberg, M.B., Eijsink, V.G.H. & Nes, I.F. Functional analysis of promoters involved in quorum sensing-based regulation of bacteriocin production in *Lactobacillus*. *Molecular Microbiology* **37**, 619-628 (2000).
67. Van Der Ploeg, J.R. Regulation of bacteriocin production in *Streptococcus mutans* by the quorum-sensing system required for development of genetic competence. *Journal of Bacteriology* **187**, 3980-3989 (2005).
68. Merritt, J., Kreth, J., Shi, W. & Qi, F. LuxS controls bacteriocin production in *Streptococcus mutans* through a novel regulatory component. *Molecular Microbiology* **57**, 960-969 (2005).
69. Qi, F., Merritt, J., Lux, R. & Shi, W. Inactivation of the *ciaH* Gene in *Streptococcus mutans* Diminishes Mutacin Production and Competence Development, Alters Sucrose-Dependent Biofilm Formation, and Reduces Stress Tolerance. *Infection and Immunity* **72**, 4895-4899 (2004).
70. de Saizieu, A. et al. Microarray-Based Identification of a Novel *Streptococcus pneumoniae* Regulon Controlled by an Autoinduced Peptide. *Journal of Bacteriology* **182**, 4696-4703 (2000).
71. Upton, M., Tagg, J., Wescombe, P. & Jenkinson, H. Intra-and Interspecies Signaling between *Streptococcus salivarius* and *Streptococcus pyogenes* Mediated by SalA and SalA1 Lantibiotic Peptides. *Journal of Bacteriology* **183**, 3931-3938 (2001).
72. Fontaine, L. et al. Quorum-sensing regulation of the production of Blp bacteriocins in *Streptococcus thermophilus*. *Journal of Bacteriology* **189**, 7195-7205 (2007).
73. Schmitz, S., Hoffmann, A., Szekat, C., Rudd, B. & Bierbaum, G. The lantibiotic mersacidin is an autoinducing peptide. *Applied and Environmental Microbiology* **72**, 7270-7277 (2006).
74. Kleerebezem, M., Kuipers, O.P., de Vos, W.M., Stiles, M.E. & Quadri, L.E.N. A two-component signal-transduction cascade in *Carnobacterium piscicola* LV17B: two signaling peptides and one sensor-transmitter. *Peptides* **22**, 1597-1601 (2001).
75. Kuipers, O.P., Beerthuyzen, M.M., de Ruyter, P.G.G.A., Luesink, E.J. & de Vos, W.M. Autoregulation of nisin biosynthesis in *Lactococcus lactis* by signal transduction. *Journal of Biological Chemistry* **270**, 27299-27304 (1995).
76. Coburn, P.S., Pillar, C.M., Jett, B.D., Haas, W. & Gilmore, M.S. *Enterococcus faecalis* senses target cells and in response expresses cytolysin. *Science's STKE* **306**, 2270 (2004).
77. Korgaonkar, A.K. & Whiteley, M. *Pseudomonas aeruginosa* enhances production of an antimicrobial in response to N-acetylglucosamine and peptidoglycan. *Journal of Bacteriology* **193**, 909-917 (2011).
78. Guerra, N. & Pastrana, L. Influence of pH drop on both nisin and pediocin production by *Lactococcus lactis* and *Pediococcus acidilactici*. *Letters in Applied Microbiology* **37**, 51-55 (2003).
79. Mortvedt-Abildgaa, C. et al. Production and pH-Dependent Bactericidal Activity of Lactocin S, a Lantibiotic from *Lactobacillus sake* L45. *Applied and Environmental Microbiology* **61**, 175-179 (1995).
80. Hindré, T., Pennec, J.P., Haras, D. & Dufour, A. Regulation of lantibiotic lactacin 481 production at the transcriptional level by acid pH. *FEMS Microbiology Letters* **231**, 291-298 (2004).
81. McAuliffe, O., O'Keeffe, T., Hill, C. & Ross, R.P. Regulation of immunity to the two-component lantibiotic, lactacin 3147, by the transcriptional repressor LtnR. *Molecular Microbiology* **39**, 982-993 (2001).

82. Franz, C.M.A.P. et al. Atypical Genetic Locus Associated with Constitutive Production of Enterocin B by *Enterococcus faecium* BFE 900. *Applied and Environmental Microbiology* **65**, 2170-2178 (1999).
83. Ihssen, J. & Egli, T. Specific growth rate and not cell density controls the general stress response in *Escherichia coli*. *Microbiology* **150**, 1637-1648 (2004).
84. Collet, A. et al. Impact of rpoS Deletion on the Proteome of *Escherichia coli* Grown Planktonically and as Biofilm. *Journal of Proteome Research* **7**, 4659-4669 (2008).